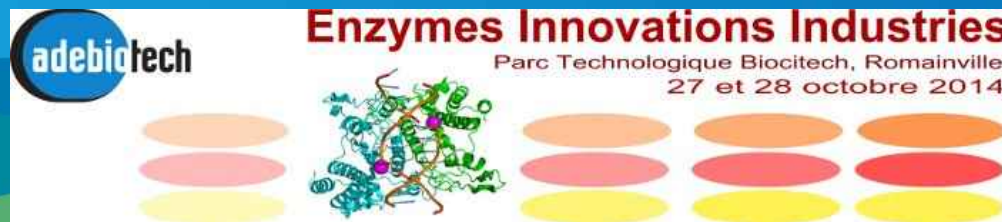


Novel enzyme development: Screening & design

Noël van Peij - Principal Scientist Strain Development & Screening
DSM Biotechnology Center, Delft, the Netherlands



Mission

Our purpose is to create brighter lives for people today and generations to come

We connect our unique competences in Life Sciences and Materials Sciences to create solutions that nourish, protect and improve performance



Building on an impressive history



Hoffman La Roche's
Vitamins (1930s)

Chemical synthesis &
biotechnology



Gist-Brocades (1869)

Biotechnology

Life Sciences



DSM (1902)

Energy, chemistry &
polymer technology

Materials Sciences

Vitamins
Omega's
Carotenoids
Premixes for food & feed
Enzymes
Minerals
Cultures & Yeasts
Nutraceuticals
Pharmaceuticals
Cellulosic bioethanol
Biomedical materials
Bio-plastics
High Performance Plastics
Polyamides and precursors
Resins for coatings and
composites
Functional Materials
Solar - advanced surfaces

DSM - key activity areas

Health

Advanced, cost-effective health and medical innovations, and healthier food and beverages, to meet the needs of a growing and ageing global population



Nutrition

World's leading producer of vitamins and nutritional ingredients meeting the growing need for more nutritious and more sustainable food and animal feed



Materials

Enabling lighter, stronger, more advanced and more sustainable performance materials

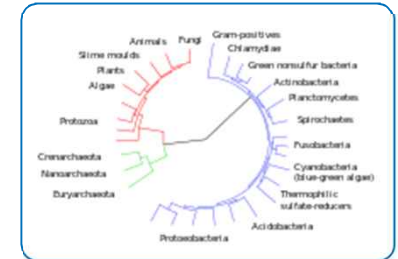


DSM's 24,500 employees deliver annual net sales of about € 10 billion

Industrial enzyme/strain development @ DSM

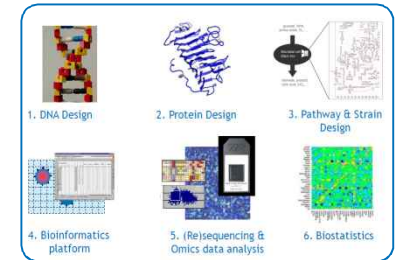
Biodiversity Platform

Enabling the identification of novel enzymes and pathways (*in-vivo*, *in-vitro* & *in-silico*), and HT (in-application) screening



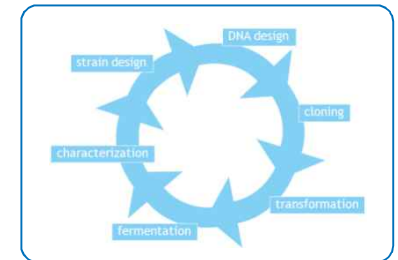
Systems Biology Platform

Enabling the modeling and design of DNA, proteins, pathways (= metabolites) being part of the cell factory in connection with experimentation



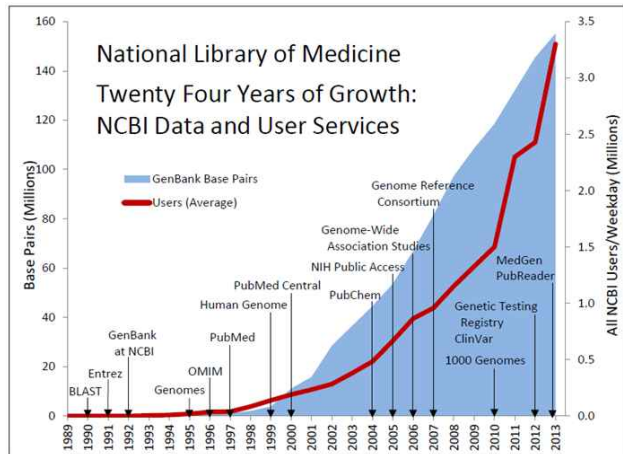
Synthetic Biology Platform

Enabling the rapid engineering & characterization of microbes, yeast, fungi and algae

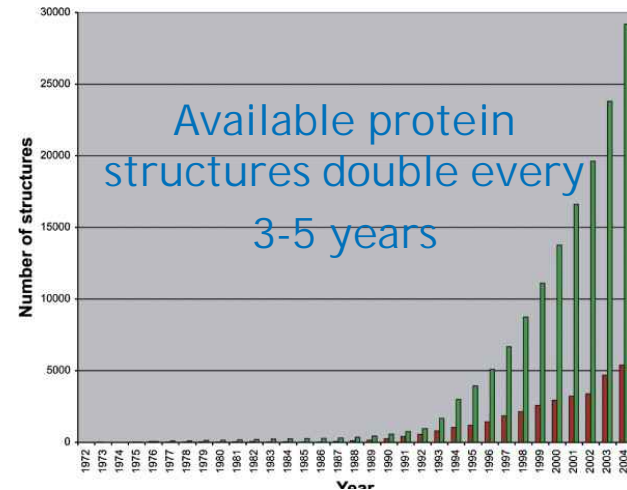


... for the production of **enzymes**, metabolites and biomass

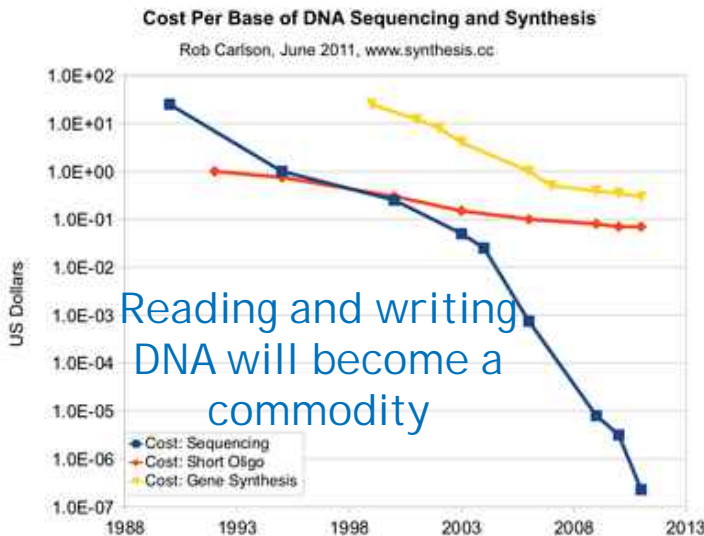
Screening and discovery: Trends



Unlimited sequence availability

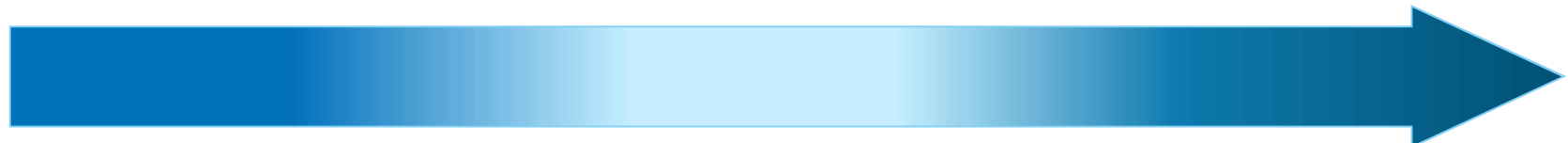


Need for standardization, parallelization & automation in strain engineering



Reading and writing DNA will become a commodity

A brief history of screening & protein engineering



1970

1980

1990

2000

2010

2020

?

Site-directed mutagenesis

Cassette/sat. mutagenesis

Error-prone PCR

DNA shuffling

smart library design

computational design?

Key driver:

- Activity-based screening

Key driver:

- Sequence-based screening
- HT screening robotics
- HT DNA sequencing

Key drivers:

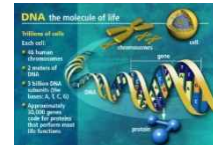
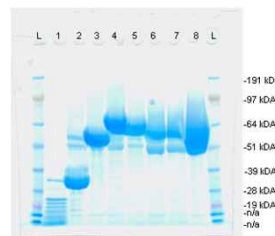
- Advanced bioinformatics & modeling
- Progressively cheaper DNA synthesis

Importance of structure information:



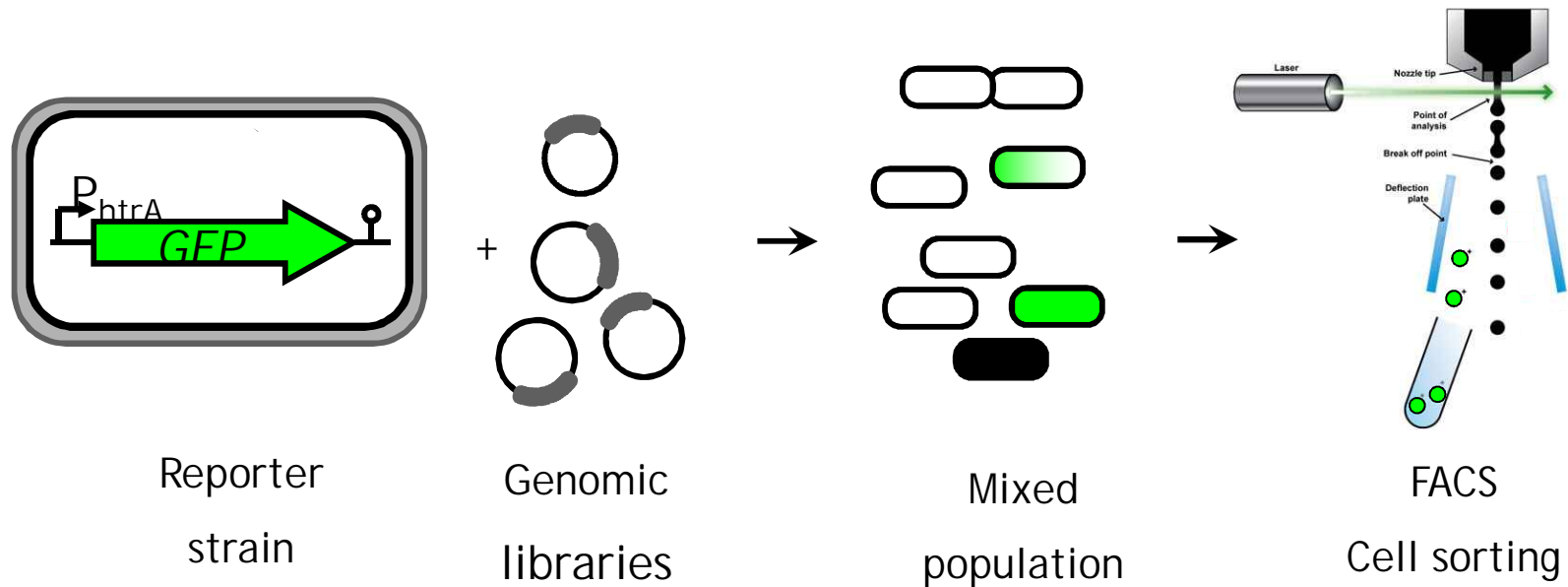
low

high



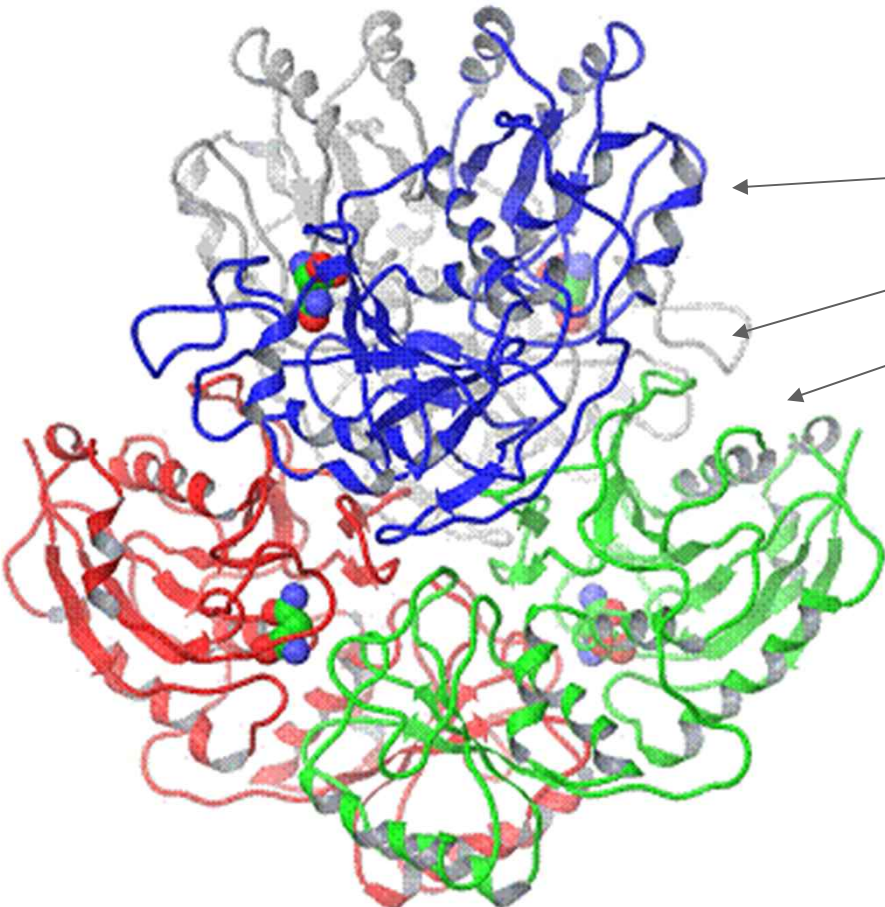
Enzyme discovery screening tools

- Strain with GFP reporter for secretion stress
- Genomic library 20.000 clones
- High GFP strains selected by FACS

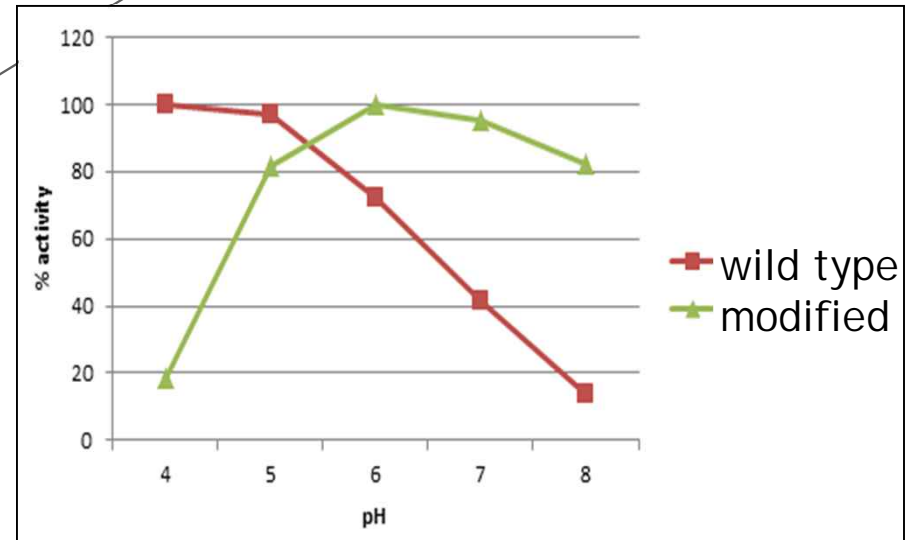


Trip (2011) Microb. Biotechnol. 4(5):673-682.

Protein engineering



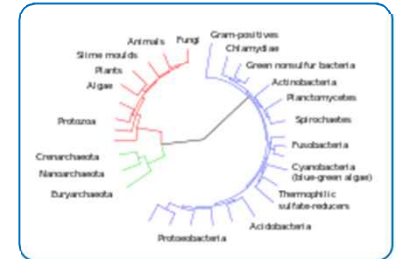
Baking enzyme example
Change Amino acid
Changes performance



Industrial strain development @ DSM

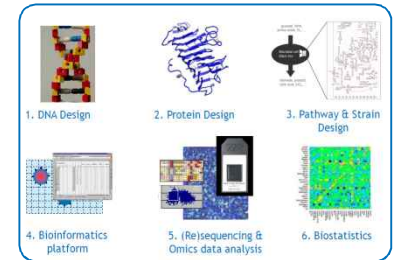
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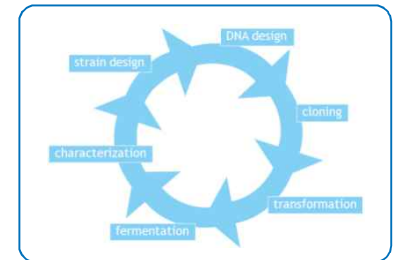
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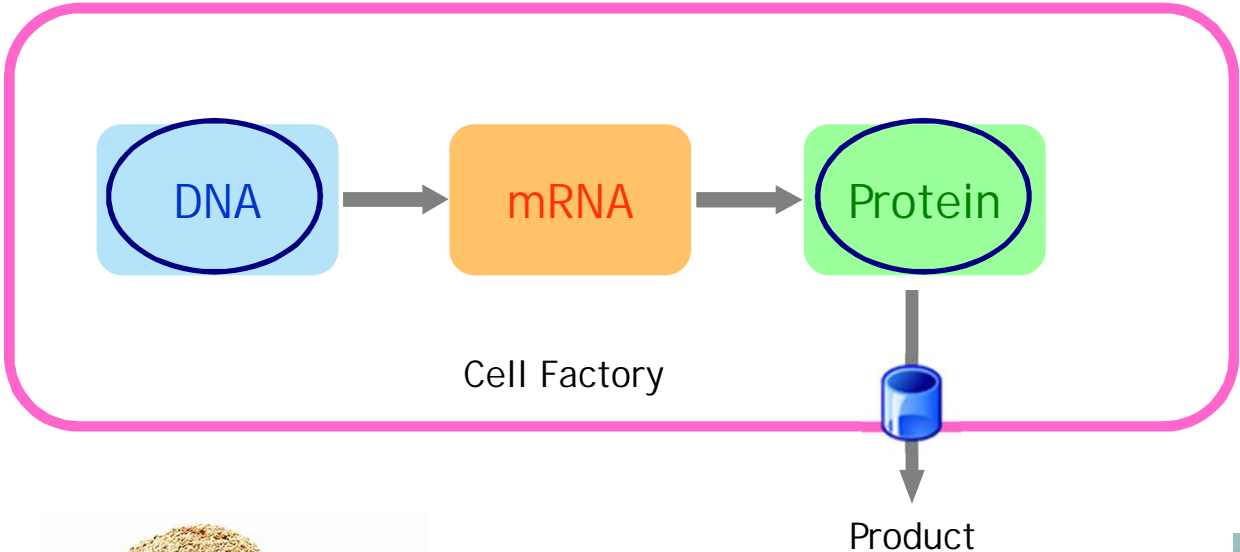
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Enabling the rapid engineering & characterization of microbes, yeast, fungi and algae



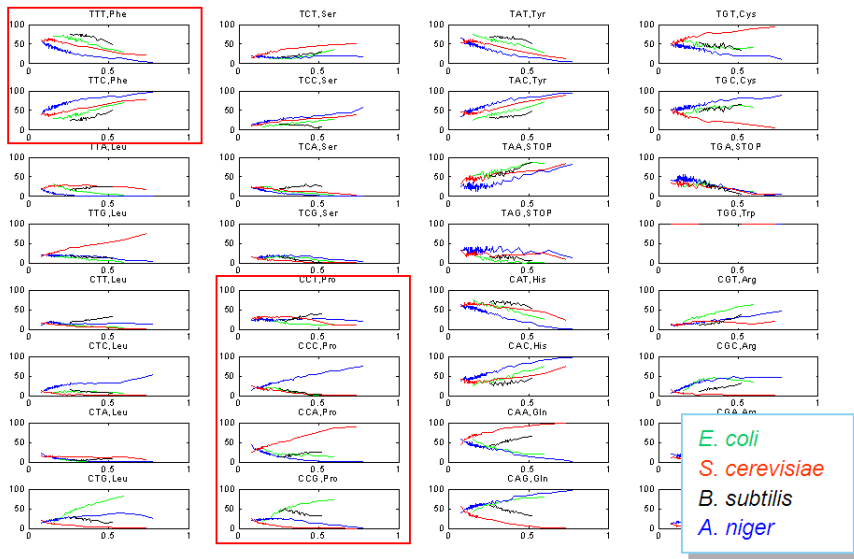
... for the production of **enzymes**, metabolites and biomass

Design of cell factory components



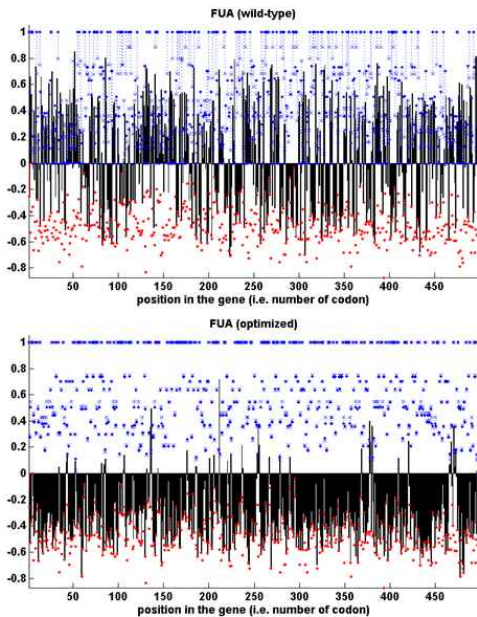
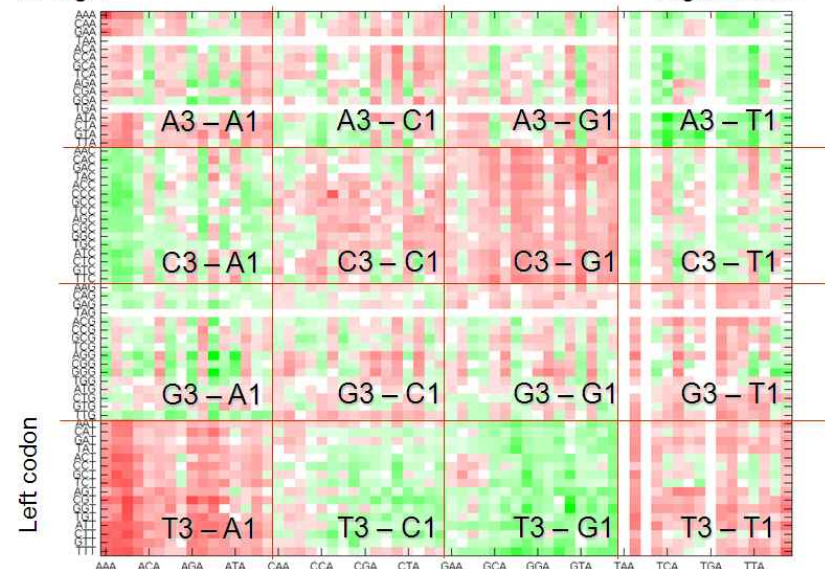
DNA sequence design to increase protein production

Green: Higher than expected based on codon statistics
 Red: Lower than expected based on codon statistics

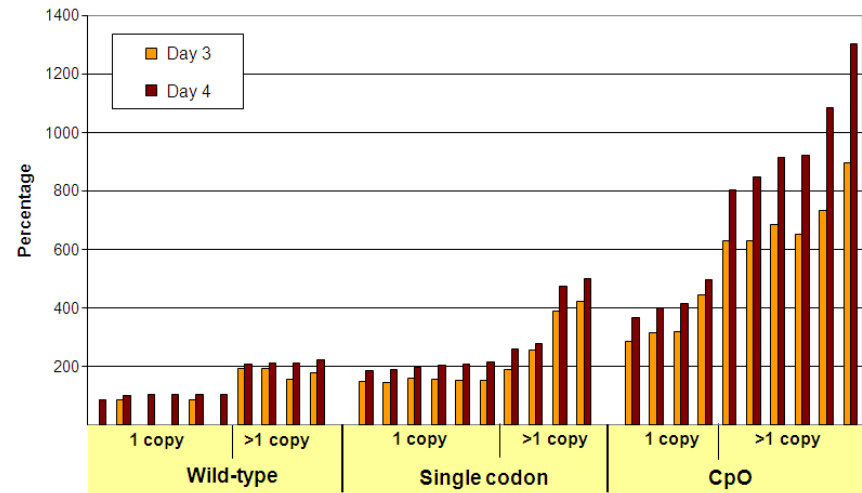
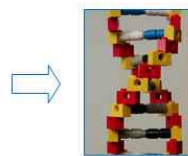


A. niger

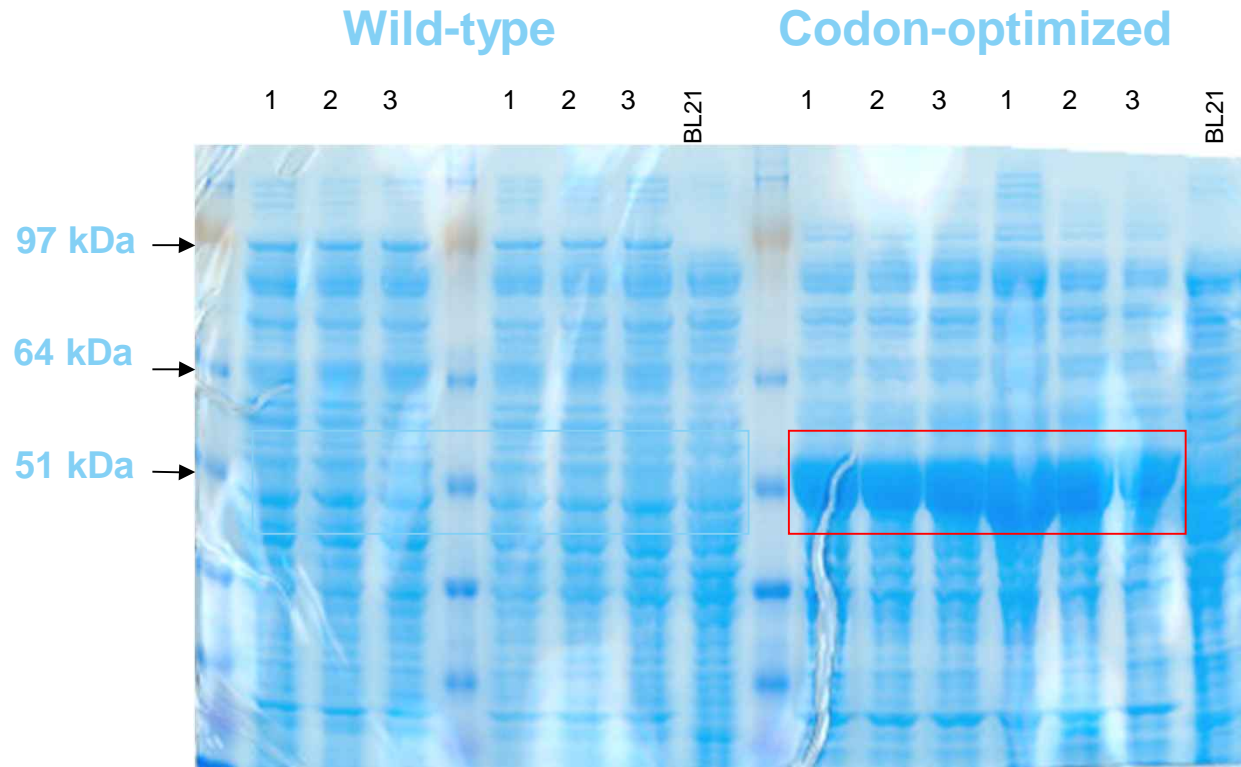
Right codon



- Statistically significant bias in codon pair usage found
- Method for optimizing genes through codon pair optimization invented and implemented
- Several optimized genes created (e.g. FUA in *A. niger*)
- WO2008/000632

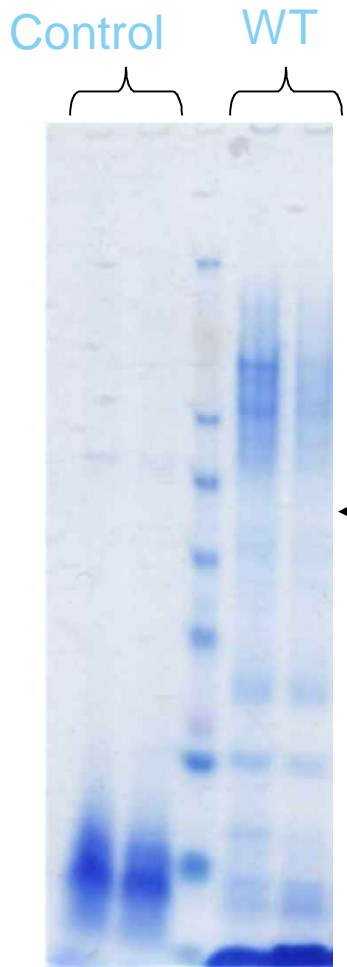


Broad application...



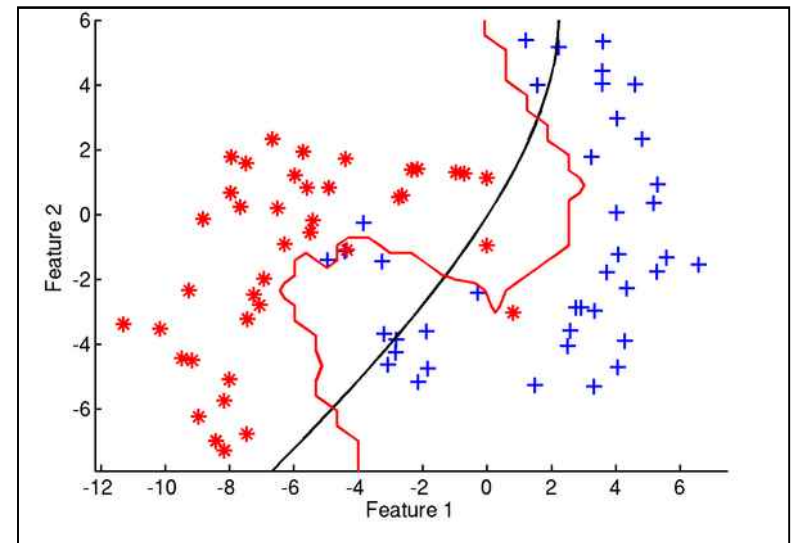
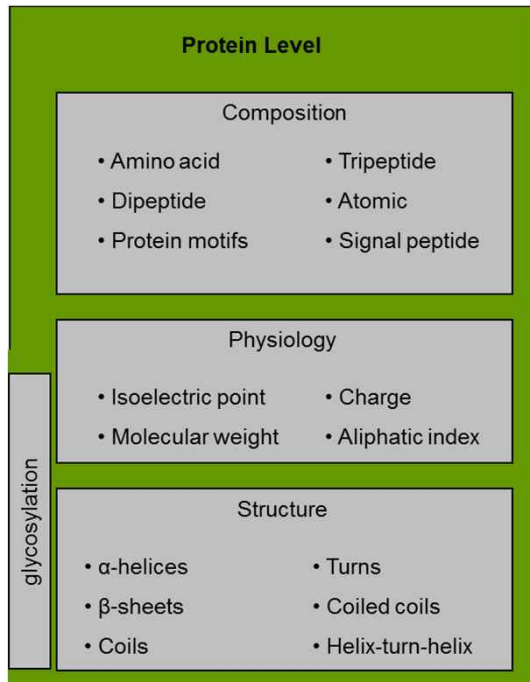
E. coli

Protein: *A. niger* chitinase



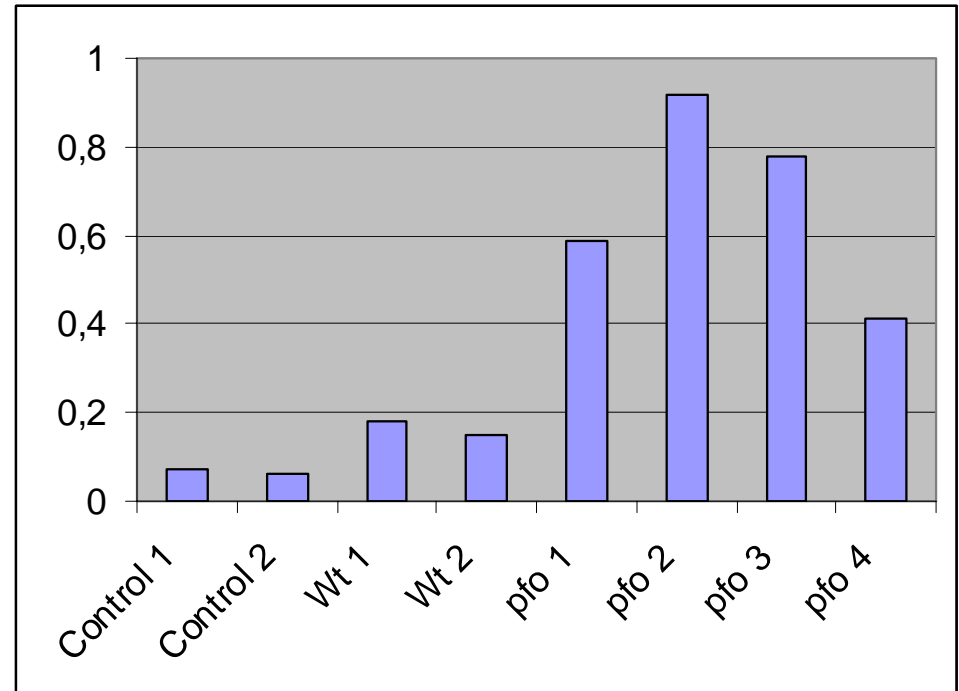
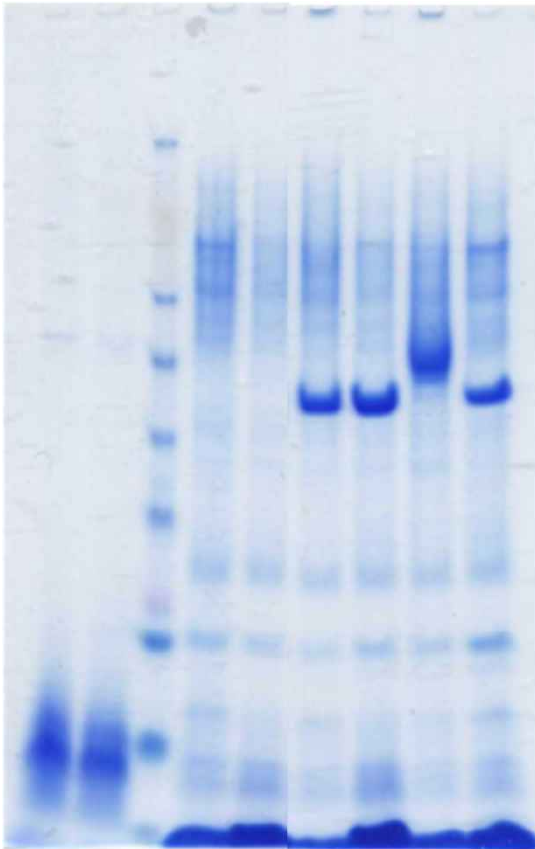
Protein: Classifier approach

- Approach:
 - Dataset of 638 overexpression attempts in *A. niger*
 - 268 successful
 - 370 not successful
 - Statistical classification to select protein features that distinguish between secreted proteins and non-secreted proteins in a dataset



Protein: *A. niger* chitinase optimized

Control M Wt Pfo1-4



WO2010/102982

Novel enzyme development: Screening & design

Target activity & matrix effect



Diversity sources



Filters: class specific, BioIT, smart design, etc



Assay strategy



Enzyme leads to PMCs

Industry needs

Enzyme class selection

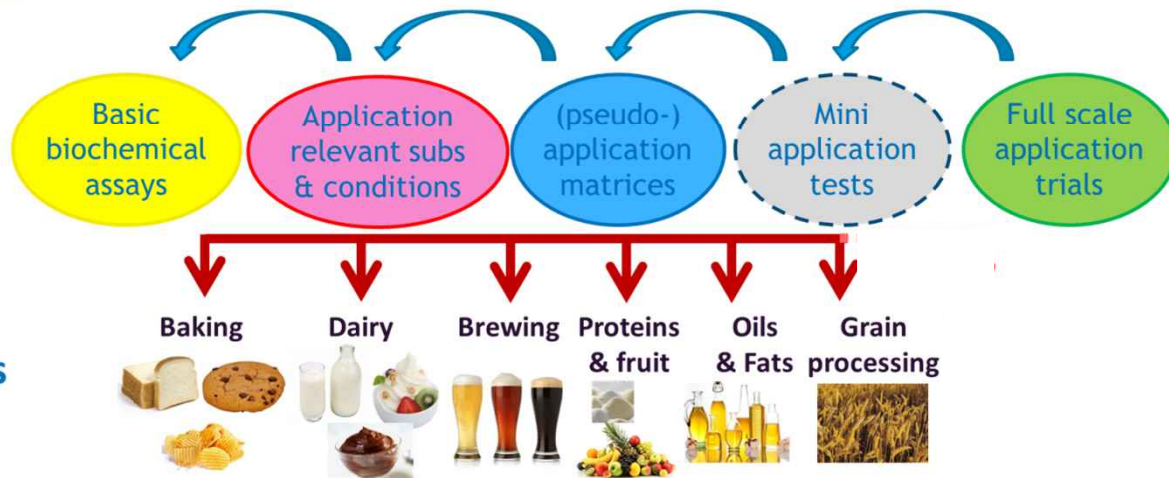


1000's

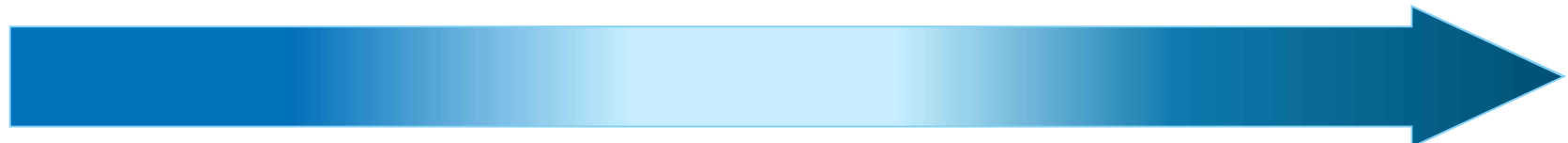
100's

10's

BioIT-driven selection and design of candidate enzymes
Historical approach
Enzyme screening
→ better focus, less screening



A brief history of screening & protein engineering



1970

1980

1990

2000

2010

2020

?

Site-directed mutagenesis

Cassette/sat. mutagenesis

Error-prone PCR

DNA shuffling

smart library design

computational design?

Key driver:

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- HT DNA sequencing

Key drivers:

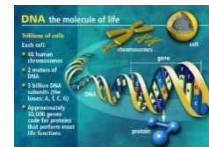
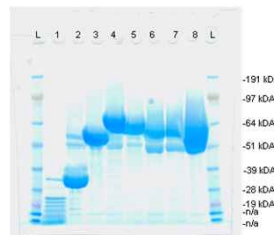
- Advanced bioinformatics & modeling
- Progressively cheaper DNA synthesis

Importance of structure information:



low

high



IMPROVE: Smart Library Design - Less = More

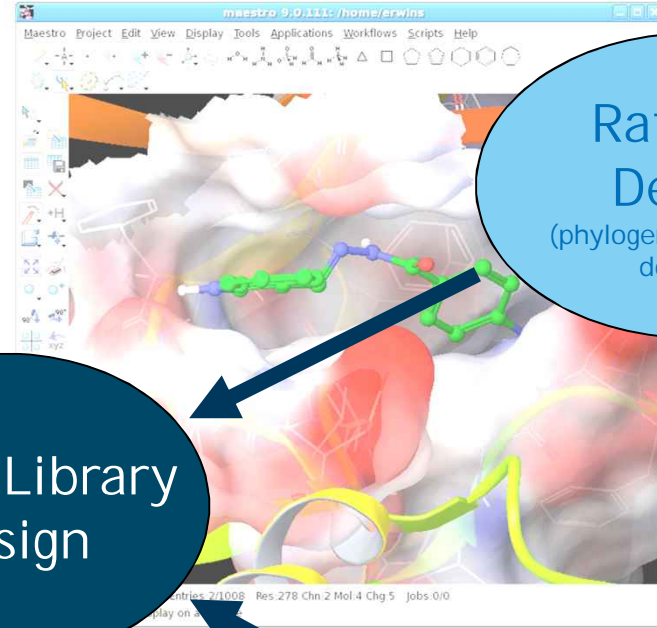
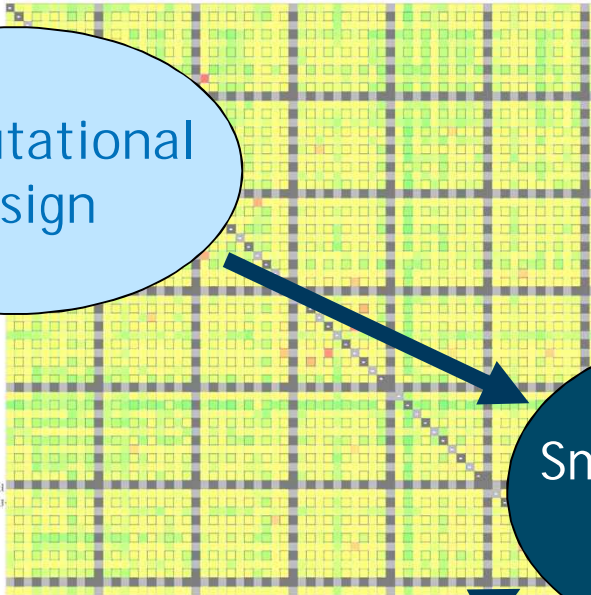
Computational Design

Rational Design
(phylogeny, structure, docking)

Smart Library Design

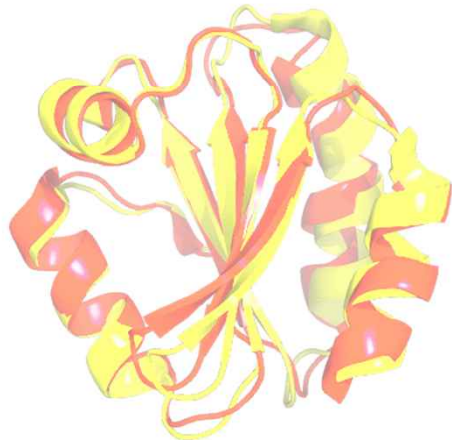
Random
(error-prone, saturation, recombination)

Statistics
(library statistics & simulation, data analysis)



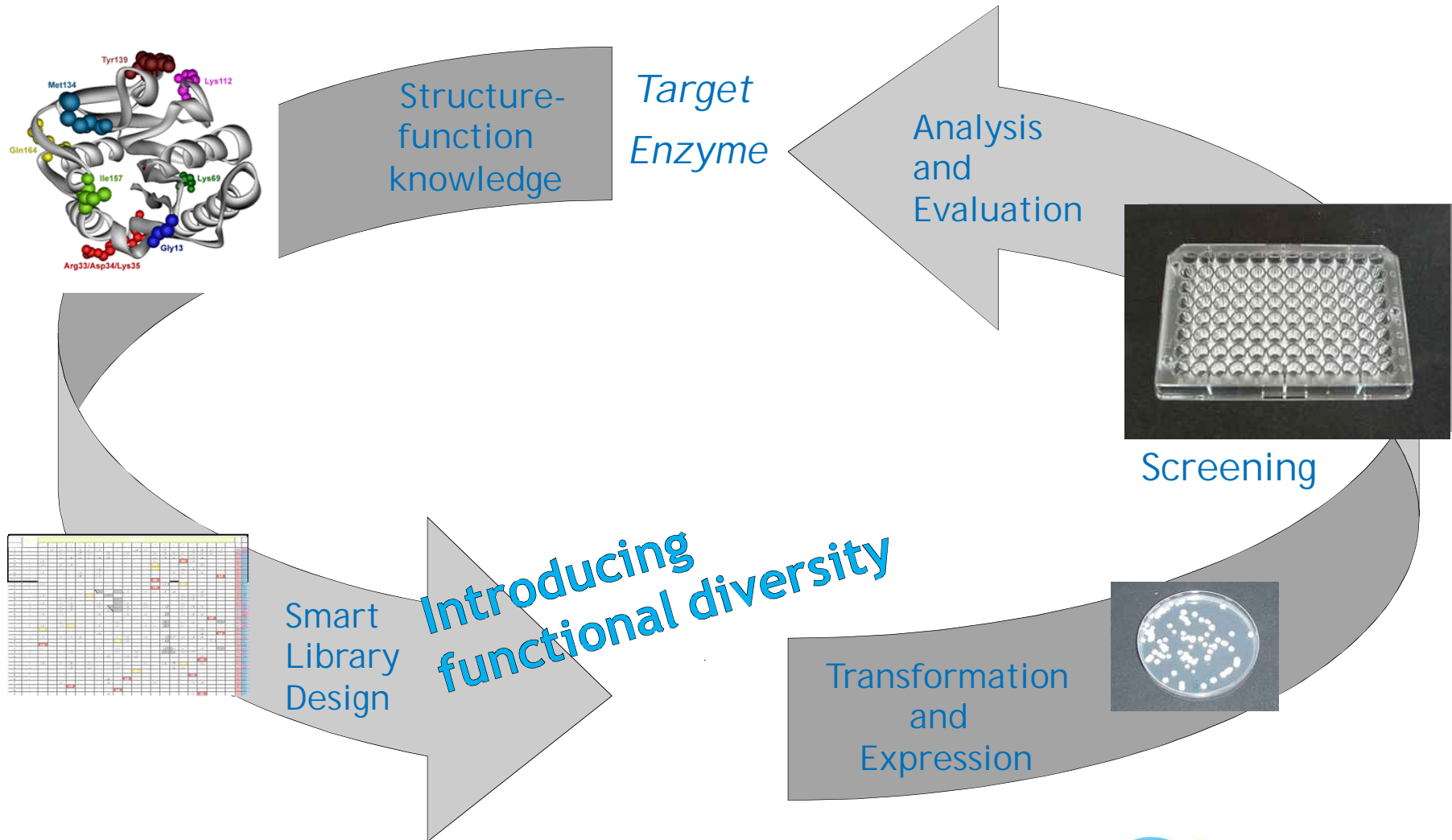
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T-COFFEE, Version 1.37 (Med
Notredame, Higgins, Hering
CPU TIME:103 sec.
SCORE=73
+
B:AD AVS GOOD
oth_OPSPD_SHEEP : 74
oth_OPSPD_BOVIN : 74
oth_OPSPD_HOUSE : 74
oth_OPSPD_HIRSH : 74
oth_OPSPD_CARAU : 71
oth_OPSPG_CARAU : 72
oth_OPSPD_XENLA : 74
oth_OPSPD_CHICK : 74
oth_OPSPD_LAMJA : 74
oth_OPSPG_CHICK : 74
oth_OPSPB_GECGE : 74
oth_OPSPU_BRARE : 74
```

```
oth_OPSPD_SHEEP : TLVIRHDKQFRHCHMITLLCCG
oth_OPSPD_BOVIN : TLVIRHDKQFRHCHVITLLCCG
oth_OPSPD_HOUSE : TLVIRHDKQFRHCHMITLLCCG
oth_OPSPD_HIRSH : TLVIRHDKQFRHCHMITLLCCG
oth_OPSPD_CARAU : TLVIRHDKQFRHCHMITLLCCG
oth_OPSPG_CARAU : TLVIRHDKQFRHCHMITLLCCG
oth_OPSPD_XENLA : TLVIRHDKQFRHCHMITLLCCG
oth_OPSPD_CHICK : TLVIRHDKQFRHCHMITLLCCG
oth_OPSPD_LAMJA : TLVIRHDKQFRHCHMITLLCCG
oth_OPSPG_CHICK : TLVIRHDKQFRHCHMITLLCCG
oth_OPSPB_GECGE : TLVIRHDKQFRHCHMITLLCCG
oth_OPSPU_BRARE : TLVIRHDKQFRHCHMITLLCCG
Cons: * * * * * : : : : : * * * * *
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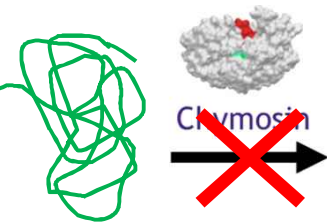
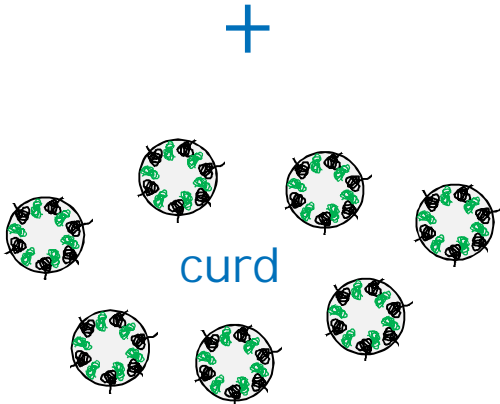
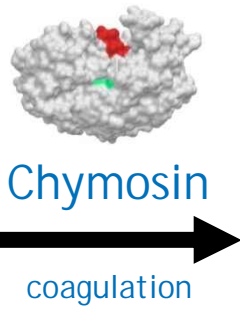
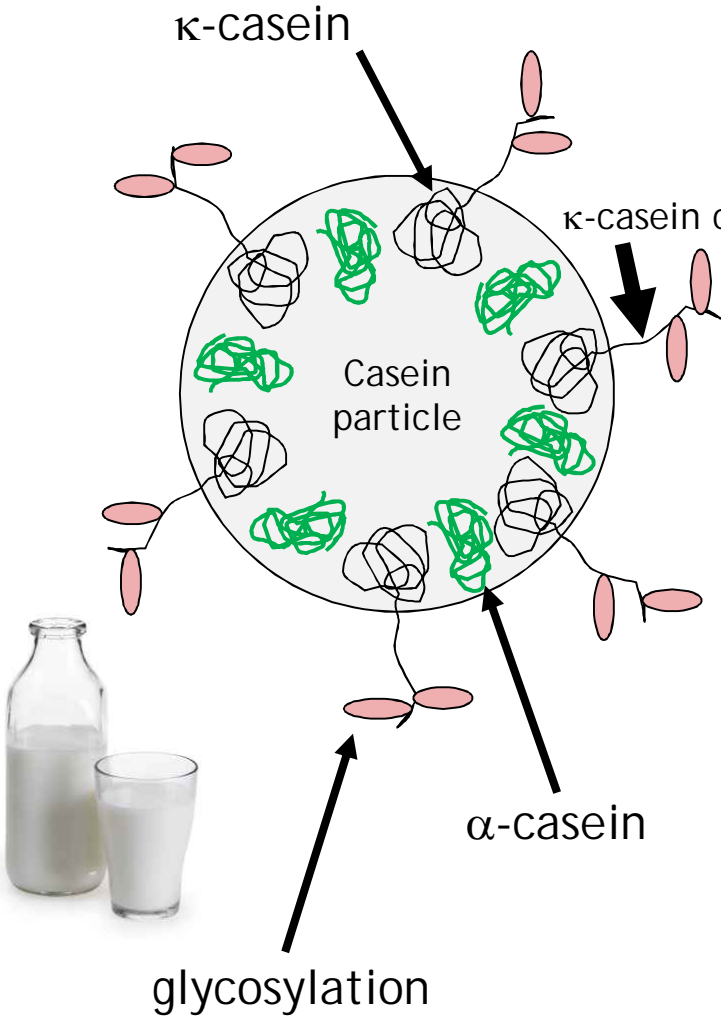
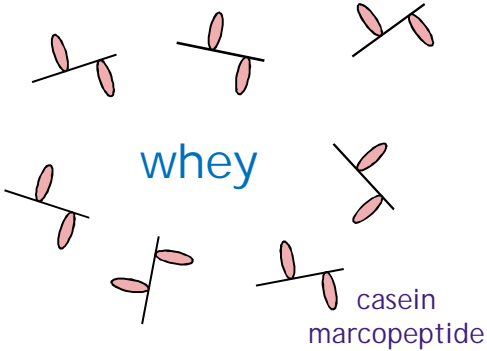


Library	Size	Seq. ID	Clust. ID	Seq. ID	Clust. ID	Seq. ID	Clust. ID	Seq. ID	Clust. ID	Seq. ID	Clust. ID	Seq. ID	Clust. ID	Seq. ID	Clust. ID	Seq. ID	Clust. ID
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2	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
3	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
4	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
5	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
6	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
7	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
8	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
9	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
10	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
11	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
12	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
13	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
14	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
15	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
16	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
17	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
18	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
19	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
20	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
21	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
22	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
23	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
24	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
25	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
26	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
27	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
28	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
29	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
30	1000	0.98	0.18	0.10	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01

The enzyme optimization cycle



The protease action of chymosin in milk



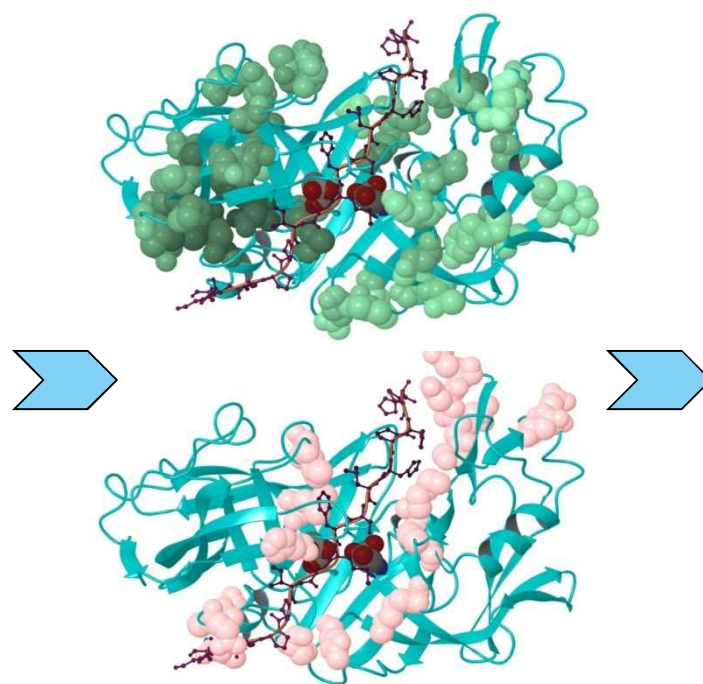
Target: limit proteolysis of α -casein (high C/P: coagulation/proteolysis ratio)



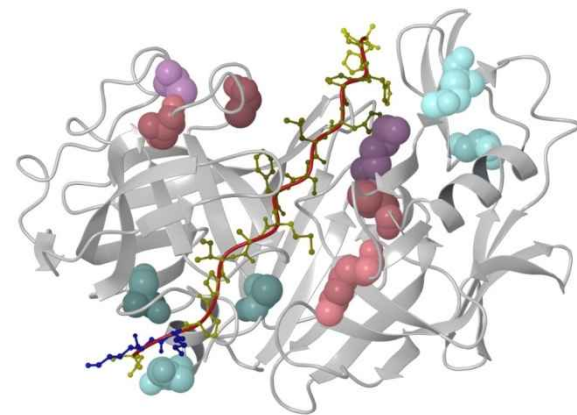
Development of high specificity chymosin



Structural analysis of mammalian chymosin sequence variations



Chymosin variants

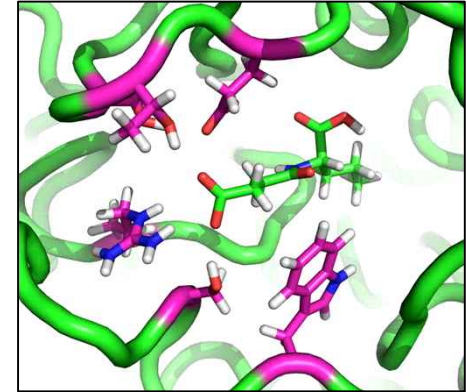
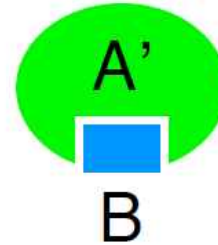
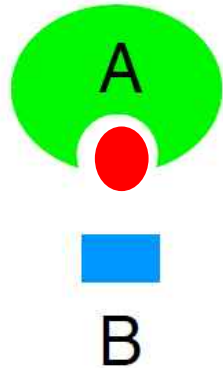


Hits with improved Specificity / activity

Unprecedented hit rates using Smart Library Design

Library type	# screened	Enzyme	Target improvement	Hit rate
random	6000	A	specific activity	0.4%
saturation	4000	A	specific activity	2.5%
saturation (sequenced)	1000	B	substrate selectivity	2.5%
Combinatorial	7000	B	substrate diversity	3%
Smart Library	85 (70% active)	Chymosin	Specificity Activity	>10%

Can we predict novel specificity sequence space *in silico*?



Input: Wild-type enzyme
and desired substrate

Output: Optimal sequences
for binding desired substrate

Advantages:

- Drastic reduction of sequence space (wrt full saturation mutagenesis)
- Reduced screening efforts (# clones & rounds)
- Combinatorial libraries allow screening for synergistic mutations

Ollikainen, de Jong, Kortemme (2014), *in preparation*

Conclusions

- Biodiversity rapidly exploding through sequencing
- Rationalization of
 - Protein expression optimization
 - Biodiversity screening
 - Enzyme engineering ('smart library')
 - Computational design...

→ efficient, faster & predictive enzyme development



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